

Product Description

Loop-mediated isothermal amplification (LAMP) is a nucleic acid amplification technology. This kit adopts the dye-based isothermal amplification method and can be used for the qualitative detection of DNA in samples (e.g., viruses). Bst2.0 HS is derived from *Bacillus stearothermophilus* DNA Polymerase I. Through genetic engineering techniques, it retains the 5' → 3' DNA polymerase activity while eliminating the 5' → 3' exonuclease activity and exhibits strand displacement capability. It is suitable for DNA strand displacement reactions and LAMP (Loop-mediated isothermal amplification). Bst2.0 HS has been genetically modified to improve its sensitivity, and compared to the wild-type Bst DNA polymerase and long fragments, this enzyme can effectively increase the amplification speed and yield. Bst2.0 HS is a thermosensitive isothermal amplification enzyme obtained via reversible modification technology based on genetically engineered Bst2.0 HS DNA polymerase. The activity of this enzyme can be completely inactivated at room temperature, allowing reactions to be initiated under ambient conditions. This suppresses non-specific amplification and improves reaction efficiency.

Components

1. 10×RT-LAMP Premix (DG) (Bst2.0 HS)
2. 5×RT-LAMP Lyophilization Buffer
3. 50×RT-LAMP Fluorescent Dye II
4. AcuGenix™ Lyophilization Protectant-9 (optional)

Storage

Store at -20±5℃. Vortex before use. Avoid freeze-thaw cycles.

Prepare Reaction Mix

Composition	Volume per Reaction	Volume per Reaction	Final Concentration
10×RT-LAMP Premix (DG) (Bst2.0 HS)	2.5 µL	5 µL	1×
5×RT-LAMP Lyophilization Buffer	5 µL	10 µL	1×
50×RT-LAMP Fluorescent Dye II	0.5 µL	1 µL	1×
4×AcuGenix™ Lyophilization Protectant-9 ¹	6.25 µL	12.5 µL	1×
10×Primer Mix ²	2.5 µL	5 µL	1×
Template RNA ³	—	—	—
ddH2O	To 25 µL	To 50 µL	—

1. This system refers to a lyophilization system. When customers operate this system without lyophilization requirements, there is an option to add the AcuGenix™ Lyophilization Protectant-9. In cases where lyophilized products are required, the AcuGenix™ Lyophilization Protectant-9 must be added during the performance validation of the product at the liquid reagent stage to ensure consistency with the components and effects of the lyophilized system.

2. Recommended primer concentration: The 10×Primer Mix comprises 16 µM FIP, 16 µM BIP, 2 µM F3, 2 µM B3, 4 µM LoopF, and 4 µM LoopB. Concentration gradient testing can be carried out to determine the optimal primer dosage. It is recommended that primers be prepared with TE Buffer.

3. Different types of RNA templates have different copy numbers of target genes. When necessary, gradient dilution can be performed to determine the optimal quantity of RNA template to be added. This product functions as a pH-sensitive indicator reagent. High-concentration Tris-HCl buffer systems should be avoided in sample elution solutions. The use of strong acid or strong alkali nucleic acid release agents for sample processing should be avoided. If such reagents are unavoidably used, it is advisable to adjust the pH to neutral.

Reaction Conditions

Incubate at a temperature range of 60-65 °C for a duration of 30-60 minutes, and acquire fluorescence data at one-minute intervals. After the reaction is completed, allow the sample to cool to ambient temperature before analyzing the results.

Prepare Reaction Mix for lyophilization

Components	25 μ L System
10 $\times$ RT-LAMP Premix (DG) (Bst2.0 HS)	2.5 μ L
5 $\times$ RT-LAMP Lyophilization Buffer	5 μ L
50 $\times$ RT-LAMP Fluorescent Dye II	0.5 μ L
AcuGenix™ Lyophilization Protectant-9	6.25 μ L
10 $\times$ Primer Mix	2.5 μ L
ddH ₂ O	To 18-20 μ L

*For other lyophilization preparation systems, please inquire separately.

Lyophilization Programe

Step	Temperature	Time	Condition	Pressure
Pre-freezing	4°C	30 min	Hold	1 atm
	-50°C	60 min	Cooling	
	-50°C	210 min	Hold	
First Drying	-30°C	60 min	Heating	Ultimate Vacuum
	-30°C	720 min	Hold	
Second Drying	25°C	60 min	Heating	Ultimate Vacuum
	25°C	300 min	Hold	

1. This freeze-drying process is an in-situ freeze-drying process for a 25 μ L system. If need to perform freeze-drying of freeze-dried beads or other in-situ systems, please inquire separately.
2. The above freeze-drying process is for reference only. Different product types and the use of different freeze dryers may result in different parameters, so adjustments can be made based on actual usage.
3. Different freeze-drying processes may be suitable for different batch sizes, and sufficient testing and verification must be conducted when used for large-scale production.

Protocol

1. Centrifuge the lyophilized sample immediately.
2. Add the nucleic acid template to the lyophilized sample and adjust the volume to 25 μ L with water.
3. Mix the sample thoroughly, centrifuge it, and load it onto the instrument.

Quality control

1. Functional testing: sensitivity, specificity, reproducibility of LAMP.
2. No exogenous nuclease activity, no exogenous endo/exonuclease contamination.

Q & A

LAMP amplification products are scarce or non-existent:

1. Elevate the quantity of the template and the concentration of the enzyme.
2. Substitute the primers. It is advisable to design multiple primer sets and conduct screening for the optimal primers via preliminary experiments. For primer design, refer to <http://primerexplorer.jp/e/>.
3. The use of Loop Primers is recommended to enhance amplification efficiency and reduce amplification time.